

H.B.O. IMPAIRS Ca⁺⁺ RESPONSES IN A CULTURED EPITHELIAL CELL LINE (HT29)

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In order to assess the effects of hyperbaric oxygen (H.B.O.) on cellular functions, we measured the Ca⁺⁺ signal pathway activation in HT29 colonic cells exposed to different doses of H.B.O. Hyperbaric oxygen exposures were carried out in a cylindrical steel chamber (40 cm diameter x 65 cm length-Galeazzi model - Italy). The chamber was constantly ventilated at a rate of 4 l/min to avoid carbon dioxide accumulation during pressurization. Gases were analyzed and O₂ concentration was > 99% (Taylor servomex OA 272 oxygen analyzer) and carbon dioxide below 0.2% (Med. Gas Analyzer LB-2, Model 24OM, Beckman).

Incubation of 10⁶ cells/ml to 4 absolute O₂ atm for 1 hour, completely prevented the cell Ca⁺⁺ increase produced by 500 μM carbachol, a muscarinic receptor agonist, meanwhile exposure to 2 absolute O₂ atm for 30 min blunted the Ca⁺⁺ response to the agonist. In fact, control cells stimulated by equimolar amounts of carbachol displayed a 515±22% increase in cytosolic Ca⁺⁺ (from 150±8 to 615±12 nM, n=4) significantly higher than that showed by cells challenged by H.B.O treatment.

In these latter, carbachol produced a 248±15% of Ca⁺⁺ increase (from 137±6 to 475 nM, n=4, P<0.001). Similar behaviour was found when cells were stimulated by thapsigargin, a microsomial Ca⁺⁺ ATPase inhibitor. Cell - staining with vital dye tripan blue revealed an elevated positive percentage (about 30-35%) of H.B.O. treated cells.

This finding shows that H.B.O. treatment impaired cell function coupled to Ca⁺⁺ signaling pathway. It is possible that this impairment, probably caused by plasma membrane damage, as suggested by tripan blue dye exclusion test, could be the molecular basis for the beneficial effect of H.B.O. therapy.